

Reversal of sulphonamide action in *Escherichia coli* (B_{12} auxotroph) by vitamin B_{12}

Sulphonamides are known to block sequentially the synthesis in *Escherichia coli* of methionine, xanthine, serine, thymine, valine^{1,2} and glycine³. At each of these steps, the inhibition index is increased in presence of vitamin B_{12} ^{2,3}. Since the requirement of vitamin B_{12} for growth of *E. coli* B_{12} auxotroph can be met completely by methionine⁴, it was of interest to study the growth inhibition by sulphanilamide (SA) in this mutant as influenced by the two metabolites.

The mutant strain⁴ of the organism was grown in the medium of GREEN AND SEVAG⁵ with additions as shown (Table I). Growth was measured after 24 hours incubation at 30° and expressed in terms of galvanometer deflections in a Klett-Summerson photoelectric colorimeter at 660 m μ .

TABLE I
COMPARATIVE ACTIVITY OF METHIONINE AND VITAMIN B_{12} IN OVERCOMING SA
GROWTH INHIBITION OF *E. coli* B_{12} AUXOTROPH

Additions to 10 ml basal medium	mg SA added			
	0	0.5	0.7	1.0
	Growth at 24 hours			
1. None	0	0	0	0
2. Methionine, 100 μ g	48	0	0	0
3. Methionine, 1 mg	48	0	0	0
4. Vitamin B_{12} , 2 m μ g	47	46	17	10
5. Vitamin B_{12} , 20 m μ g	49	48	39	23
6. As in (3) + xanthine 0.25 mg	48	30	17	14
7. As in (6) + serine 0.2 mg	50	49	36	19

It is observed (Table I) that, in presence of vitamin B_{12} , a higher concentration of SA is required to inhibit growth than when methionine is used. The protective effect of vitamin B_{12} is more pronounced at the higher concentration of the vitamin and can be simulated by a combination of methionine, xanthine and serine.

These observations suggest that, although methionine could, interchangeably with vitamin B_{12} , serve the requirement for growth of the mutant, this vitamin assumes additional functions in a condition of stress caused by SA bacteriostasis analogous to its *p*-aminobenzoic acid potentiating action already observed² with the wild strain under similar conditions. That the requirements for growth and for metabolic activity with respect to specific systems need not necessarily be the same is known⁶.

H. R. ALIMCHANDANI
A. SREENIVASAN

Department of Chemical Technology, University of Bombay (India)

¹ K. C. WINKLER AND P. G. DE HAAN, *Arch. Biochem.*, 18 (1948) 97.

² W. SHIVE, *Ann. N.Y. Acad. Sci.*, 52 (1950) 1212.

³ H. R. ALIMCHANDANI AND A. SREENIVASAN, *Nature* (communicated).

⁴ B. D. DAVIS AND E. S. MINGIOLI, *J. Bacteriol.*, 60 (1950) 17.

⁵ M. N. GREEN AND M. G. SEVAG, *Arch. Biochem.*, 9 (1946) 129.

⁶ E. E. SNELL, in *Bacterial Physiology*, edited by C. H. WERKMAN AND P. W. WILSON, Academic Press, Inc., New York, 1951, pp. 215-255; W. D. BELLAMY AND I. C. GUNSALUS, *J. Bacteriol.*, 48 (1944) 191.

Received September 17th, 1955

Relationship between skew diffusion gradient curves and axial ratios of rod-shaped particles

It is a well-known phenomenon that from experiments with filamentous molecules skew diffusion gradient curves are obtained, in contrast to the symmetrical curves obtained from those with spherical particles. Thus POLSON¹, GRALÉN² and JULLANDER³ found that in plotting diffusion curves of cellulose derivatives, which are known to be filamentous, the curves obtained are asymmetrical. KAHLER⁴ made similar observations in his diffusion experiments on solutions of sodium thymonucleate, and NEURATH AND SAUM⁵ obtained skew diffusion curves in their work